

# 22/6/2020

To – Nano Z coating Shedlovski 1 st. YAVNE 8122101

## Microbiological Test -

<u>Chemical disinfectants and antiseptics - Quantitative carrier test</u>

<u>For the evaluation of bactericidal and fungicidal activity for instruments used in the medical and food areas</u>

**Laboratory Number: 20055427/1-3** 

Sample description: CPD - ALCO STERILE

Date sample received: 2/6/2020

Date Tested: 17/6/2020

### 1. Standard:

The test was conducted based on Israeli Standard 1944, BS EN 14561 "Evaluation of bactericidal activity" and AAMI TIR 12 (2010).

## **Test Purpose:**

This test was conducted in order to define the antimicrobial effectiveness of the disinfectant preparation (CPD – ALCO STERILE).

## Inoculation:

- 1.1 Stainless steel surfaces (4/4cm) were sterilized by steam.
- 1.2 The surfaces were inoculated with the following bacteria (four surfaces for each microorganism) –

Staphylococcus aureu	IS ATCC 6538	
Pseodomonas aerugii	nosa ATCC 9027	
Escherichia coli	ATCC 8739	
Aspergillus niger	ATCC 16404	
Enterococcus faecalis	ATCC 51299	
Lactobacillus plantrar	rum ATCC 14917	
Salmonella typhimuri	um ATCC 14028	

Page 1 of 5

#### Remarks

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- 3. The results are related only to the tested sample.
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Enterobacter aerogenes ATCC 13048	
Saccharomyces cerevisiae ATCC 51299	
Listeria monocytogenes ATCC 19115	
Bacillus atrophaeus spores ATCC 9372	

The bacterial suspensions were diluted using soil (ATS - containing Proteins Healthmark (MI, US)) to give a final concentration of bacteria Of  $10^5-10^7$  per/surface (about 100 µl from each suspension was added To the soil according to cell turbidity).

- 1.3 One surface was not inoculated negative control.
- 1.4 The surfaces were left to dry in biohazard hood for 30 minutes.

## 2. Test Procedure:

- 2.1 The tests are divided to three parts –
- 2.1.1 55427-1 The surfaces were inoculated and left for 24 hours in the laminar hood and then were sprayed with the tested sample and were tested after drying of the surfaces for the efficacy of the disinfection.
- 2.1.2 55427-2 The surfaces were sprayed and left for 5 hours in the laminar hood and then were contaminated and were tested after drying of the surfaces for the efficacy of the disinfection.
- 2.1.3 55427-3 The surfaces were sprayed and left for 10 hours in the laminar hood and then were contaminated and were tested after drying of the surfaces for the efficacy of the disinfection.
- 2.2 Two surfaces before disinfection and cleaning for each microorganism were placed aseptically into sterile cups. 100 ml were added to each sample (Neutralizing solution lot 904) and vortexed for 1 minute and then the diluted sample was plated according to the pour plate technique using warm TSA (lot 16570) or SDA (lot 16528) or APT (lot 16620).
- 2.3 The plates were incubated for 72 hours at 30-35°C or 120 hours at 20-25 °C for yeasts and moulds. After incubation of the test plates, the Microorganisms were counted on each plate.
- 2.4 The remaining surfaces (two for each microorganism) were subjected to disinfection according to manufacturer's instructions (contact for 15 and 60 seconds according to manufactures instructions). One surface after disinfection was put into cups and 0.1 ml of Neutralizing solution

Page 2 of 5

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- was spread on each surface. The surfaces were then diluted with 10 ml (BPS+1% Tween 80 lot 16358) and vortexed for  $\underline{1}$  minute.
- 2.5 Then the eluent was plated in the pour plate technique using TSA or SDA or APT. One surface was touched using rodac plates by (TSA +Lec +Polys 80)
- 2.6 The plates were incubated as defined in 2.1 and then the microbial count was determined per surface.

## 3. Results:

Bacteria/Yeast/ Mould	55427-1 Before disinfection CFU/surface	55427-1 After disinfection CFU/surface	55427-2 Before disinfection CFU/surface	55427-2 After disinfection CFU/surface
<i>P. aeruginosa</i> ATCC 9027	186,000,000	640	8,400,000	280
	180,000,000	860	9,200,000	320
S.aureus ATCC 6358	102,000,000	260	9,600,000	260
	95,000,000	280	10,200,000	280
E.coli ATCC 8739	102,000,000	<10	11,200,000	220
	98,000,000	1	9,200,000	240
Aspergillus niger	4,800,000	<10	2,600,000	140
ATCC 16404	5,000,000	1	3,200,000	180
Enterococcus faecalis	136,000,000	<10	9,600,000	360
ATCC 51299	140,000,000	<1	11,400,000	380
Lactobacillus	4,800,000	60	5,200.000	220
plantrarum ATCC 14917	5,000,000	40	4,200,000	190
Salmonella typhimurium ATCC 14028	136,000,000 140,000,000	<10 2	11,600,000 10,600,000	260 240
Enterobacter aerogenes ATCC 13048	126,000,000	960	11,600,000	320
	95,000,000	1080	10,200,000	440
Saccharomyces	112,000,000	20	9,800,000	260
cerevisiae ATCC 51299	110,000,000	30	9,200,000	320

Page 3 of 5

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<i>Listeria monocytogenes</i>	206,000,000	10	9,200,000	320
ATCC 19115	180,000,000	20	9,600,000	240
Candida albicans ATCC	102,000,000	<10	4,200,000	180
10231	110,000,000	<1	3,400,000	140
NC	<10		<10	

Bacteria/Yeast/ Mould	55427-3 Before disinfection CFU/surface	55427-3 After disinfection CFU/surface
P. aeruginosa ATCC 9027	18,600,000 19,200,000	220 260
S.aureus ATCC 6358	6,800,000 7,200,000	320 240
E.coli ATCC 8739	15,400,000 16,800,000	180 260
Aspergillus niger ATCC 16404	470,000 580,000	320 460
Enterococcus faecalis ATCC 51299	14,600,000 16,400,000	460 320
Lactobacillus plantrarum ATCC 14917	960,000 880,000	240 180
Salmonella typhimurium ATCC 14028	18,800,000 20,200,000	380 320
Enterobacter aerogenes ATCC 13048	18,200,000 17,800,000	280 320
Saccharomyces cerevisiae ATCC 51299	14,600,000 15,400,000	360 420
Listeria monocytogenes ATCC 19115	18,600,000 19,800,000	380 420
Candida albicans ATCC 10231	3,600,000 3,200,000	160 280
NC	<10	

Page 4 of 5

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### 4. Conclusion:

- 4.1 According to the test results, the disinfection using CPD in the presence of organic soil was able to reduce 5-6 magnitudes (at least 99.999%) for the microorganism tested in the first option (immediate disinfection).
- 4.2 According to the test results, the disinfection using CPD in the presence of organic soil was able to reduce 3-5 magnitudes (at least 99.9%) for the microorganism tested in the second option (contamination 5 hours after disinfection).
- 4.3 According to the test results, the disinfection using CPD in the presence of organic soil was able to reduce 3-5 magnitudes (at least 99.9%) for the microorganism tested in the third option (contamination 10 hours after disinfection).

Authorized Signature Avraham PhD Microbial Labratory

Preformed by:\_

Page 5 of 5

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